

Phytochemical Studies on *Amberboa ramosa*

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ABSTRACT

Twelve compounds have been isolated for the first time from *Amberboa ramosa* namely, octacosanoic acid (**1**), cinnamic acid (**2**), 7,8-dihydroxycoumarin (**3**), 5-hydroxy-7,8-dimethoxyflavone(**4**), 5-hydroxy-3,6,7,4'-tetramethoxyflavone (**5**), 5,7,3',4'-tetrahydroxyflavone (**6**), β -amyrin (**7**), β -amyrin acetate (**8**), marrusidin A (**9**) marrusidin B (**10**), polyodonine (**11**), stigmasterol 3-*O*- β -D-glucopyranoside (**12**)respectively. Their structures have been elucidated by EI-MS, HR-EI-MS, HR-FAB-MS, ¹H-NMR and ¹³C-NMR spectroscopic data. Compounds **2-6** showed moderate antioxidant activity.

Keywords: NMR; *Amberboa ramosa*; Compositeae; Antioxidant activity; Compositeae.

1. INTRODUCTION

The genus *Amberboa* is a small genus of family Compositeae, comprising six species. These are annuals or biennials extending from Mediterranean to Central Asia¹⁻². This plant is used as tonic, aperient, febrifuge, deobstruent, cytotoxic and antibacterial activities. A decoction of leaves is given to relieve swelling and to dissolve kidney stones, treating vomiting, paralysis and diseases of spleen. A paste of this plant is used on open wounds and other skin diseases. Its roots and flowers are reported to possess medicinal properties against rheumatism and kidney troubles³⁻⁵. Shifting of literature revealed that triterpenoids, flavonoids, steroids and sesquiterpene lactones have previously been reported from this species⁶⁻⁸. The chemotaxonomic and ethnopharmacological significance of the genus *Amberboa* prompted us to reinvestigate the constituents of *A. ramosa*. As a result, we now report the isolation and structural elucidation of octacosanoic acid (**1**), cinnamic acid (**2**), 7,8-dihydroxycoumarin (**3**), 5-hydroxy-7,8-dimethoxyflavone(**4**), 5-hydroxy-3,6,7,4'-tetramethoxyflavone (**5**), 5,7,3',4'-tetrahydroxyflavone (**6**), β -amyrin (**7**), β -amyrin acetate (**8**), marrusidin A (**9**) marrusidin B (**10**), polyodonine (**11**), stigmasterol 3-*O*- β -D-glucopyranoside (**12**), respectively. The compounds **2-6** showed moderate antioxidant activity.

2. RESULTS AND DISCUSSION

The methanolic extract of the whole plant of *Amberboa ramosa* was subsequently divided into *n*-hexane, chloroform, ethylacetate, *n*-butanol and water-soluble fractions. The ethyl acetate soluble fraction was subjected to a series of column and flash chromatographic techniques as described in the experimental to obtain twelve compounds reported for the first time from this species. These could be identified octacosanoic acid (**1**), cinnamic acid (**2**), 7,8-dihydroxycoumarin (**3**), 5-hydroxy-7,8-dimethoxyflavone(**4**), 5-hydroxy-3,6,7,4'-tetramethoxyflavone (**5**), 5,7,3',4'-tetrahydroxyflavone (**6**), β -amyrin (**7**), β -amyrin acetate (**8**), marrusidin A (**9**) marrusidin B (**10**), polyodonine (**11**), stigmasterol 3-*O*- β -D-glucopyranoside (**12**) respectively, on the basis of their respective spectral data.

3. EXPERIMENTAL

3.1 General

Optical rotations were measured on a JASCO DIP-360 polarimeter. IR spectra were recorded on a 460 Shimadzu spectrometer. EI-MS and HR-FAB-MS were recorded on JMS-HX-110 and JMS-DA 5000 mass spectrometers. The ¹H-NMR, ¹³C-NMR, HMQC, and HMBC spectra were recorded on Bruker spectrometers operating at 400 MHz for ¹H- and 100 MHz for ¹³C-NMR, respectively. The chemical shift values are reported in ppm (δ) units and the coupling constants (J) are in Hz. Aluminum sheets precoated with silica gel 60 F₂₅₄ (20 × 20 cm, 0.2 mm thick; E-Merck) were used for TLC and silica gel (230-400 mesh) was used for column chromatography. Visualization of the TLC plates was carried out under UV at 254 and 366 nm and by spraying with ceric sulfate reagent (with heating). Melting points were determined on a Gallenkemp apparatus and are uncorrected. For antioxidant assay all the chemicals were purchased from Sigma Chemical Company (St. Louis, MO, USA).

3.2 Plant Material

The whole plant of *Amberboa ramosa* Jafri (Compositae) was collected in June 2002, from Karachi (Pakistan) and identified by Dr. SurraiyaKhatoon, Plant Taxonomist, Department of Botany, University of Karachi, where a voucher specimen (no. KU 312 b) has been deposited.

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Received 5th January 2012, Accepted 20th January 2012

3.3 Extraction and Isolation

The shade dried plant material (22 kg) was extracted with methanol (3x40 L) for ten days at room temperature. The solvent was evaporated under reduced pressure and the residue (217 g) was partitioned between *n*-hexane and water. The water soluble fraction was further extracted with chloroform, ethyl acetate and *n*-butanol. The column chromatography of the ethyl acetate soluble fraction (90 g) over silica gel and elution with *n*-hexane-ethyl acetate in increasing order of polarity afforded six major fractions (A-F). The fraction A which was obtained from *n*-hexane: CHCl₃ (8:2) was further purified by column chromatography over silica gel eluting successively with hexane and chloroform (8.6:1.4) to afford **1** and (7.5:2.5) to furnished compound **2**. The fraction B obtained from *n*-hexane-CHCl₃ (7.0:3.0) was a mixture of three components, which were separated by column chromatography using solvent system *n*-hexane-CHCl₃ (7.7:2.3) to afford compound **4**, *n*-hexane-CHCl₃ (7.4:2.6) to afford compounds **7** and *n*-hexane-CHCl₃ (7.2:2.8) compound **8**, respectively. The fraction C obtained from *n*-hexane-CHCl₃ (5.0:5.0) was further purified by column chromatography eluting with *n*-hexane-CHCl₃ (5.6:4.4) to afford compound **3** and mixture of two compounds which was further purified by column chromatography using solvent *n*-hexane-CHCl₃ (4.9:5.1) as eluent to obtain compounds **5** and **11**. The fraction D obtained from *n*-hexane-CHCl₃ (2.0:8.0) was rechromatographed over silica gel and eluted with *n*-hexane-CHCl₃ (2.3:7.7) afford compounds **9** and **10** from the top and tail fractions, respectively. The fraction E obtained from CHCl₃ was rechromatographed and eluted with CHCl₃-MeOH (9.9:0.1) to afford compound **6**. The fraction F obtained from CHCl₃-MeOH (9.5:0.5) was subjected to column chromatography eluting with CHCl₃-MeOH (9.8:0.2) to obtain compound **12**.

3.3.1 Octacosanoic acid

Amorphous solid (21 mg); 135-136 °C; IR (KBr) ν_{max} cm⁻¹: 2780 and 1705; ¹H-NMR (CDCl₃, 400 MHz) δ: 2.32 (2H, t, *J*= 7.5 Hz, H-2), 1.61 (2H, q, H-3), 1.23-1.28 (48H, br s, H-4-H27) and 0.84 (3H, t, *J*= 6.4 Hz, Me-28); EI-MS *m/z* (rel. int.): 424 (M⁺), 368 (80), 325 (9), 269 (16), 185 (14), 111 (31), 101 (13), 87 (23), 73 (100), 60 (55); HR-EI-MS *m/z*: 424.4276 (calcd for C₂₈H₅₆O₂, 424.4280). The physical and spectral data showed complete agreement with those reported in the literature⁹.

3.3.2 Cinnamic acid

White solid (8 mg), m.p. 132-133 °C; UV (CHCl₃) λ_{max} (log ε) nm: 326 (3.9), 285 (3.06); ¹H-NMR (CDCl₃, 400 MHz) δ: 7.52 (1H, d, *J* = 15.9 Hz, H-1'), 7.36 (m), 7.29 (m), 7.21 (m), 6.25 (1H, d, *J* = 15.9 Hz, H-2'); HR-EI-MS, *m/z*: 148.6151 (calcd for C₉H₈O₂, 148.6158), EI-MS *m/z*(rel. int.): 148 [M]⁺ (21), 117 (25), 107 (100), 105 (16). The physical and spectral data showed complete agreement with those reported in the literature¹⁰.

3.3.3 7,8-Dihydroxycoumarin

Amorphous solid (14 mg), m.p. 113-114°C; UV (CHCl₃) λ_{max} (log ε) nm: 312 (3.77), 243 (3.82), 218 (4.08); IR (KBr) ν_{max} cm⁻¹: 3108, 1713, 1607, 1595, 1525, 1503; ¹H-NMR (CDCl₃, 400 MHz) δ: 7.61 (1H, d, *J* = 9.5 Hz, H-4), 6.85 (1H, d, *J* = 8.4 Hz, H-5), 6.75 (1H, d, *J* = 8.4 Hz, H-6), 6.10 (1H, d, *J* = 9.5 Hz, H-3); HR-EI-MS, *m/z*: 178.0261 (calcd for C₉H₆O₄, 178.0267), EI-MS *m/z*(rel. int.): 178 (100), 150 (84), 122 (14), 94 (28), 66 (43), 51 (14). The physical and spectral data were much closed to the reported values¹¹.

3.3.4 5-Hydroxy-7,8-dimethoxyflavone

Yellowish crystalline solid (18 mg); M.P.:97-98 °C; UV (MeOH) λ_{max} nm (log ε): 247 (3.28), 215 (3.50), 315 (4.52) IR(KBr) ν_{max} cm⁻¹:3519, 2925, 1660, 1610, 1570; ¹H-NMR(CD₃OD, 400 MHz) δ: 6.68 (s, H-6), 7.85 (1H, d, *J* = 7.6 Hz, H-2', H-6'), 7.53 (3H, m, H-3', H-4', H-5'), 6.58 (1H, s, H-3), 3.92, 3.98 (6H, s, MeO-7, 8) and 12.69 (chelated hydroxyl group at C-5); EI-MS*m/z* (rel. int.): 298 (37), 255 (28), 181 (15), 153 (53), 125 (17), 102 (19), 93 (21), 77 (25), 69 (100); HR-EI-MS: [M]⁺ peak atm/*z*: 298.0829 (calcd for C₁₇H₁₄O₅; 298.0831). The physical and spectral data were in closed agreement to the reported values¹².

3.3.5 5-Hydroxy-3,6,7,4'-tetramethoxyflavone

Amorphous powder (12 mg), m.p. 178-180 °C; UV (CHCl₃) λ_{max} (log ε) nm: 331 (3.92), 275 (3.89), 252 (4.06); IR (KBr) ν_{max} cm⁻¹: 3502, 2954, 1672, 1621, 1564, 1509; ¹H-NMR (CDCl₃, 400 MHz) δ : 8.05 (2H, d, *J* = 8.5 Hz, H-2', H-6'), 6.99 (2H, d, *J* = 8.5, H-3', H-5'), 6.48 (1H, s, H-8), 3.93, 3.90, 3.87, 3.84 (12H, s, MeO-6, 7, 3, 4'), and 12.60 due to chelated hydroxyl group at C-5; HR-EI-MS showed [M]⁺ at *m/z* 358.1123 (calcd for C₁₉H₁₈O₇, 358.1129), E-IMS *m/z* (rel. int.): 358 (100), 344 (57), 316 (11), 300 (18), 226 (20), 165 (16), 135 (28), 83 (25). The physical and spectral data identified it as 5-hydroxy-3,6,7,4'-tetramethoxyflavone¹³⁻¹⁴.

3.3.6 5,7,3',4'-Tetrahydroxyflavone

Yellow crystals (10 mg), m.p. 328°C; UV (CHCl₃) λ_{max} (log ε) nm: 256 (3.64), 265 (3.62), 360 (4.12) and 306 (4.41); IR (KBr) ν_{max} cm⁻¹: 3300 (OH), 1655 (C=O) and 1593 (C=C); ¹H-NMR (C₅D₅N, 400 MHz) δ: 6.72 (1H, d, *J* = 1.8 Hz, H-6), 6.91 (1H, d, *J* = 1.8 Hz, H-8), 6.79 (1H, s, H-3), 7.54 (1H, dd, *J* = 8.2, 2.2 Hz, H-6'), 7.27 (1H, d, *J* = 8.2 Hz, H-5'), 7.51 (1H, d, *J* = 2.2 Hz, H-2'), 13.78 (1H, br s, 5-OH); ¹³C-NMR (C₅D₅N, 100 MHz) δ : 181.7 (C-4), 166.9 (C-7), 157.9 (C-5), 101.8 (C-9), 162.6 (C-2) , 114.7 (C-3), 150.4 (C-3'), 151.9 (C-4'), 147.8 (C-6'), 116.8 (C-1'), 119.4 (C-

5'), 122.7 (C-2'), 164.0 (C-10); HR-EI-MS, *m/z*:286.2321 (calcd for C₁₅H₁₀O₆, 286.2365), EI-MS *m/z* (rel. int.): 286 (100), 270 (34), 153 (16) and 69 (18). The physical and spectral data showed complete agreement with those reported in literature¹⁵.

3.3.7 β -Amyrin

Crystallized from ethanol (33 mg); MP: 197-198 °C; [α]_D²⁵: +100(*c* = 0.21, CHCl₃); IR (KBr) ν_{max} cm⁻¹: 3510, 3055, 1635 and 820; ¹H-NMR (CDCl₃, 500 MHz) δ: 5.11 (1H, m, H-12), 3.19 (1H, dd, *J* = 10.0, 4.5 Hz, H-3), 1.02, 1.01, 1.08, 0.96, 0.93, 0.88, 0.85 and 0.80 (3H, each s, Me); EI-MS *m/z* (rel. int.): 426 [M]⁺ (15), 411 (18), 408 (16), 393 (32), 257 (20), 218 (100), 207 (10), 203 (40) and 189 (55); HR-EI-MS *m/z*: 426.3825 (calcd. for C₃₀H₅₀O, 426.3861). The physical and spectral data were in close agreement to the reported values¹⁶⁻¹⁷.

3.3.8 β -Amyrin acetate

Crystallized from MeOH (34 mg); MP: 244-245 °C; [α]_D²⁵: +81.4(*c* = 0.20, CHCl₃); IR (KBr) ν_{max} cm⁻¹: 3055, 1710, 1660, 1460, 1382, 1180 and 810; ¹H-NMR (CDCl₃, 500 MHz) δ: 5.10 (1H, m, H-12), 4.05 (1H, dd, *J* = 10.0, 4.5 Hz, H-3), 2.13 (3H, s, OAc), 1.00, 1.07, 0.98, 0.96 (3H, each s, Me), 0.90 (6H, s, H-29 and H-30), 0.86 and 0.81 (3H, each s, Me); EI-MS *m/z* (rel. int.): 468 [M]⁺ (27), 426 (35), 411 (11), 408 (12), 257 (26), 218 (100), 207 (8), 203 (55) and 189 (75); HR-EI-MS *m/z*: 468.3931 (calcd. for C₃₂H₅₂O₃, 468.3916). The physical and spectral data coincided with the literature¹⁸.

3.3.9 Marrusidin A

Amorphous powder (38 mg); M.p. 228-230°C; [α]_D²⁵ = -42.2(*c* = 0.045, CHCl₃); IR (KBr) ν_{max} cm⁻¹: 2975, 1782, 1765, 1475, 1180 and 1050; ¹H-NMR (CDCl₃, 400 MHz) δ: 1.51-1.53 (1H, m, H-1a), 1.29-1.30 (1H, m, H-1b), 1.78-1.80 (1H, m, H-2a), 1.71-1.73 (1H, m, H-2b), 2.10-2.13 (1H, m, H-3a), 1.42-1.44 (1H, m, H-3b), 2.05 (1H, d, *J*=4.5 Hz, H-5), 4.68 (1H, t, *J*=5.4 Hz, H-6), 2.15-2.17 (1H, m, H-7a), 1.75-1.77 (1H, m, H-7b), 2.14-2.15 (1H, m, H-8), 2.07-2.08 (1H, m, H-11a), 1.86-1.87 (1H, m, H-11b), 2.37-2.39 (1H, m, H-12a), 2.17-2.19 (1H, m, H-12b), 2.41 (1H, dd, *J*=13.4, 5.5, H-14a), 2.21-2.24 (1H, m, H-14b), 5.32 (1H, dd, *J*=9.2, 5.7, H-15), 1.10 (3H, d, *J*=6.3 Hz, H-17), 1.27 (3H, s, H-18), 1.04 (3H, s, H-20), 3.45 (3H, s, OMe at C-15); EI-MS: *m/z* 378 (34), 335 (10), 320 (7), 211 (100); HR-EI-MS: *m/z*378.1340 (C₂₁H₃₀O₆; calc. 378.1346). Its physical and spectral data showed complete agreement with those reported in literature¹⁹.

3.3.10 Marrusidin B

Amorphous powder (38 mg); M.p.230-232°; [α]_D²⁵ = +21.0 (*c* 0.045, CHCl₃); IR (KBr) ν_{max} cm⁻¹: 2974, 1780, 1768, 1473, 1178 and 1055; ¹H-NMR (CDCl₃, 400 MHz) δ: 1.48-1.50 (1H, m, H-1a), 1.30-1.32 (1H, m, H-1b), 1.76-1.77 (1H, m, H-2a), 1.66-1.69 (1H, m, H-2b), 2.05-2.06 (1H, m, H-3a), 1.40-1.43 (1H, m, H-3b), 2.14 (1H, d, *J*=4.5 Hz, H-5), 4.67 (1H, t, *J*=5.2 Hz, H-6), 2.21-2.23 (1H, m, H-7a), 1.70-1.74 (1H, m, H-7b), 2.09-2.2.11 (1H, m, H-8), 2.24-2.25 (1H, m, H-11a), 1.87-1.88 (1H, m, H-11b), 2.33-2.55 (1H, m, H-12a), 2.15-2.17 (1H, m, H-12b), 2.55 (1H, dd, *J*=13.6, 5.6, H-14a), 2.18-2.19 (1H, m, H-14b), 5.36 (1H, dd, *J*=9.0, 5.8, H-15), 0.84 (3H, d, *J*=6.0 Hz, H-17), 1.25 (3H, s, H-18), 1.02 (3H, s, H-20), 3.48 (3H, s, OMe at C-15); EI-MS: *m/z* 378 (36), 335 (9), 320 (6), 211 (100); HR-EI-MS: *m/z*378.1340 (C₂₁H₃₀O₆; calc. 378.1346). The physical and spectral data showed complete resemblance with the reported values¹⁹.

3.3.11 Polyodonine

White crystalline solid (38 mg); M.p 200-201°C; [α]_D²⁵ = +78° (*c* 0.1, CHCl₃); IR (KBr) ν_{max} cm⁻¹:1760 cm⁻¹ and 1740, 3092 and 1602 cm⁻¹; ¹H-NMR (CDCl₃, 400 MHz): δ: 1.71(m, 1H), 1.31(m, 1H), 1.59(m, 1H), 1.83(m, 1H), 2.17(m, 1H), 1.52 (m, 1H), 2.28 (d, 4.4 Hz, 1H), 4.75 (dd, 4.4, 6.0 Hz), 1.73 (m, 1H), 2.23 (m, 1H), 2.15 (m, 1H), 2.37 (d, 18.4 Hz, 1H), 2.80 (d, 18.4 Hz, 1H), 4.99 (d, 2.5 Hz, 1H), 6.62 (dd, 2.5, 0.5 Hz, 1H), 4.33 (d, 10.7 Hz, 1H), 4.47 (dd, 10.7, 0.5 Hz, 1H), 0.72 (d, 6.4 Hz, 3H), 1.32 (s, 3H), 1.07 (s, 3H); ¹³C-NMR (CDCl₃, 100 MHz) δ: 28.3 (C-1), 17.7 (C-2), 28.1 (C-3), 44.0(C-4), 44.3 (C-5), 75.5 (C-6), 31.1 (C-7), 31.3 (C-3), 84.6 (C-9), 38.5 (C-10), 40.0 (C-11), 212.5 (C-12), 92.7 (C-13), 102.7 (C-14), 151.4 (C-15), 77.9 (C-16), 15.6 (C-17), 22.5 (C-18), 183.4 (C-19), 23.7 (C-20); EIMS: *m/z* (rel. intensity): 346.2 (13), 329 (6), 234 (47), 207 (3) and 173 (39). The physical and spectral data showed complete agreement with literature²⁰.

3.3.12 Stigmasteryl 3-O- β -D-glucopyranoside

Colorless crystals (38 mg); MP: 289-290 °C; [α]_D²⁵:-51.5 (*c* = 0.22, CH₃OH); IR (KBr) ν_{max} cm⁻¹: 3454 (OH), 3024, 1646 (C=C); ¹H-NMR (CD₃OD, 400 MHz) δ: 5.23 (1H, br d, *J* = 5.4 Hz, H-6), 5.14 (1H, dd, *J* = 15.2, 8.4 Hz, H-22), 5.02 (1H, dd, *J* = 15.2, 8.6 Hz, H-23), 4.78 (1H, d, *J* = 7.4Hz, H-1'), 3.83 (1H, m, H-3), 3.84-4.44 (m, Glc-H'), 1.01 (3H, s, Me-19), 0.90.(3H, d, *J* = 6.2 Hz, Me-21), 0.83 (3H, d, *J* = 6.5 Hz, Me-26), 0.82 (3H, t, *J* = 7.0 Hz, Me-29), 0.80 (3H, d, *J* = 6.5 Hz, Me-27), 0.67 (3H, s, Me-18); ¹³C-NMR (CD₃OD, 125 MHz) δ: 141.5 (C-5), 138.9 (C-22), 129.1 (C-23), 121.1 (C-6), 102.8 (C-1'), 79.8 (C-3), 76.9 (C-3'), 76.7 (C-5'), 74.2 (C-2'), 70.6 (C-4'), 62.2 (C-6'), 57.0 (C-14),

56.1 (C-17), 52.1 (C-24), 50.8 (C-9), 43.9 (C-4), 43.1 (C-13), 40.5 (C-20), 39.9 (C-12), 37.8 (C-1), 36.9 (C-10), 32.9 (C-25), 32.8 (C2), 31.9 (C-7), 31.7 (C-8), 28.9 (C-16), 25.6 (C-28), 24.5 (C-15), 21.9 (C-21), 21.7 (C-27), 21.5 (C-11), 19.5 (C-19), 19.1 (C-26), 12.6 (C-18), 12.1 (C-29); EI-MS m/z (rel. int.): 412 [M-Glu]⁺ (72), 397 (15), 394 (22), 379 (28), 369 (35), 351 (71), 300 (67), 327 (55), 301 (15), 273 (21), 271 (26); HR-FAB-MS m/z : 575.4231 [M+H]⁺ (calcd for C₃₅H₅₉O₆, 575.4233). It was identified through physical and spectral data as stigmasterol 3-O- β -D-glucopyranoside²¹⁻²².

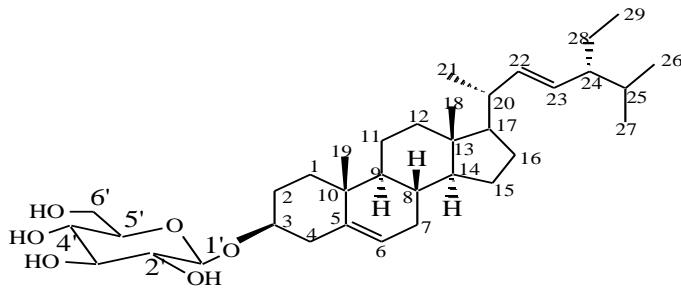
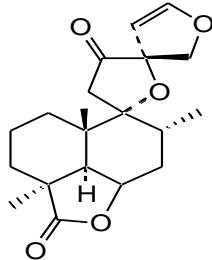
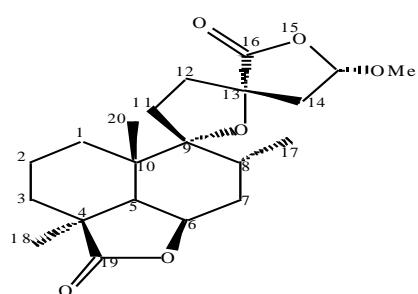
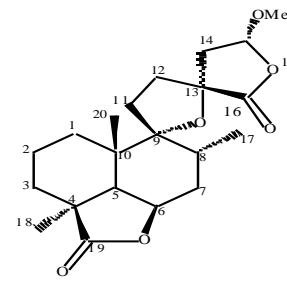
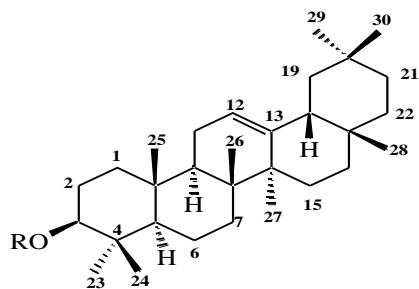
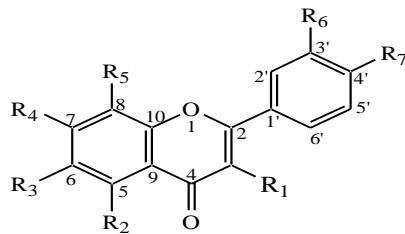
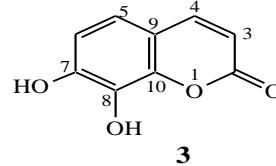
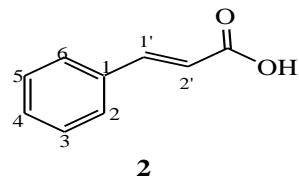
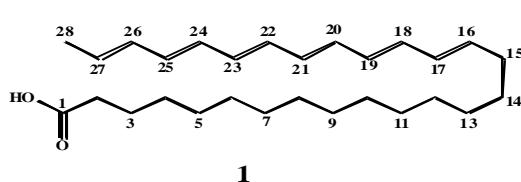


Fig-1: Structure of compounds 1-12 isolated from *Amberboa ramosa*

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